

EFFECTS OF ENZYME INHIBITORS
IN INHIBITING THE GROWTH AND
INDUCING THE DIFFERENTIATION
OF HUMAN PROMYELOCYTIC
LEUKEMIA CELLS, HL-60

Sir:

It is well known that bestatin binds to cells, most strongly to macrophages¹⁾ and enhances immune responses²⁻⁴⁾. It has also been reported that the addition of bestatin treatment to chemotherapy prolonged the survival periods of patients of nonlymphocytic leukemia⁵⁾, melanoma⁶⁾ *etc.* Recently OKUYAMA⁷⁾ reported that bestatin caused a differentiation of cancer cells. Therefore, it became necessary to clarify whether the action causing differentiation is involved in the therapeutic effect of bestatin. Besides bestatin, arphamenines A and B⁸⁾, inhibiting aminopeptidase B, amastatin⁹⁾, inhibiting aminopeptidase A, forphenicine¹⁰⁾, inhibiting alkaline phosphatase and forphenicinol¹¹⁾ (a derivative of forphenicine) have been found in this institute to be immuno-modifiers. Therefore, we examined their action in inhibiting the growth and inducing the differentiation of human promyelocytic leukemia cells, HL-60¹²⁾.

HL-60 cells were cultured in RPMI1640 medium supplemented with 15% of fetal calf serum (Grand Island Biochemical Co., Grand Island, N.Y.) and gentamicin (80 $\mu\text{g}/\text{ml}$) at 37°C in an atmosphere of 5% CO₂ in air in humidified incubators. The HL-60 cells were obtained from Dr. HOZUMI (Saitama Cancer Center Research Institute). As the marker of cell differentiation to granulocytes, intracellular blue-black formazan deposits (BFD) produced by reduction of nitro blue tetrazolium (NBT)¹³⁾ was employed. After culturing with the test materials described above for 5 days, the cells were washed with phosphate-buffered saline and incubated in RPMI1640 containing 0.1% NBT and 100 ng/ml phorbol 12-myristate 13-acetate, for 20 minutes at 37°C. After staining with May-Grünwald-Giemsa, the percentage of cells containing BFD was counted under light microscopy; at least 200 cells were examined for each experiment. The concentrations of the test materials except forphenicine were 12.5, 25, 50 and 100 $\mu\text{g}/\text{ml}$. The concentrations of forphenicine were 1.25, 2.5, 5 and 10 $\mu\text{g}/\text{ml}$. The growth was examined by Coulter Counter (Coulter Electronics Ltd.,

Table 1. Effect on growth and differentiation in HL-60 cells by proteinase inhibitors.

Drugs	Concentration ($\mu\text{g}/\text{ml}$)	% of control cells	% of BFD containing cells
Arphamenine A	100	19.1	59.0
	50	51.3	18.7
	25	76.1	9.1
	12.5	87.1	4.9
Arphamenine B	100	43.4	33.6
	50	58.2	15.8
	25	71.8	10.2
	12.5	84.3	2.3
Bestatin	100	38.3	24.6
	50	47.3	13.7
	25	50.3	9.5
	12.5	58.6	7.8
Amastatin	100	91.3	1.3
	50	97.7	1.8
	25	104.0	2.8
	12.5	107.0	1.3
Control	0	100.0	1.7

HL-60 cells were cultured at various concentration of drugs in medium for 5 days.

Table 2. Effect on growth and differentiation in HL-60 cells by enzyme inhibitors.

Drugs	Concentration ($\mu\text{g}/\text{ml}$)	% of control cells	% of BFD containing cells
Forphenicine	10	23.3	49.2
	5	73.7	3.0
	2.5	98.5	4.1
	1.25	98.6	3.3
Forphenicinol	100	102.0	1.4
	50	102.0	2.4
	25	107.0	1.1
	12.5	108.9	0.6
Control	0	100.0	1.0

England). The results are shown in Tables 1 and 2.

Arphamenine A at 100 $\mu\text{g}/\text{ml}$ showed a strong growth inhibition, and at this concentration 59.0% cells contained BFD. Arphamenine B showed weaker activity both in growth inhibition and in differentiation induction than does arphamenine A. Bestatin had further weaker activities in both actions. Amastatin at 100 $\mu\text{g}/\text{ml}$ had almost no activity. Forphenicinol at 100 $\mu\text{g}/\text{ml}$ had also almost no activity. Forphenicine was the strongest in both activities, because its 10 $\mu\text{g}/\text{ml}$ produced 77% growth in-

hibition and 49.2% differentiation induction. As the whole, it can be said that the growth inhibition always occurred in the condition where the differentiation was observed.

The time courses of the cell growth and percentage of cells containing BFD during 7 days after the addition of arphamenine A at 100 $\mu\text{g}/\text{ml}$ was examined. In this case also, the increase of the percentage of differentiated cells was accompanied by the inhibition of cell growth.

Acknowledgments

This research was partially supported by a Grant-in-Aid for Cancer Research from the Ministry of Education, Science and Culture, Japan and from the Ministry of Health and Welfare, Japan.

KEIKO MIURA
TSUTOMU SAWA
TOMIO TAKEUCHI
HAMAO UMEZAWA

Institute of Microbial Chemistry,
3-14-23 Kamiosaki, Shinagawa-ku,
Tokyo 141, Japan

(Received January 22, 1986)

References

- MÜLLER, W. E. G.; D. K. SCHUSTER, R. K. ZAHN, A. MAIDHOF, G. LEYHAUSEN, D. FALKE, R. KOREN & H. UMEZAWA: Properties and specificity of binding sites for the immunomodulator bestatin on the surface of mammalian cells. *Int. J. Immunopharmacol.* 4: 393~400, 1982
- ISHIZUKA, M.; J. SATO, Y. SUGIYAMA, T. TAKEUCHI & H. UMEZAWA: Mitogenic effect of bestatin on lymphocytes. *J. Antibiotics* 33: 653~662, 1980
- UMEZAWA, H. *Ed.*: Small Molecular Immunomodifiers of Microbial Origin. *Fundamental and Clinical Studies of Bestatin*. Jpn. Sci. Soc. Press, Pergamon Press, Tokyo, 1981
- ISHIZUKA, M.; T. TAKEUCHI, H. UMEZAWA, F. ABE, K. SHIBUYA, K. EBIHARA & T. YAMASHITA: Effect of bestatin on immune system and experimental animal tumors. *In Recent Results of Bestatin 1985. A Biological Response Modifier*. *Ed.*, H. UMEZAWA, pp. 1~12, Japan Antibiotics Res. Assoc., Tokyo, 1985
- YAMADA, K.: Immunotherapy with bestatin for acute nonlymphocytic leukemia in adults. *In Recent Results of Bestatin 1985. A Biological Response Modifier*. *Ed.*, H. UMEZAWA, pp. 73~78, Japan Antibiotics Res. Assoc., Tokyo, 1985
- IKEDA, S.; K. ISHIHARA & G. TAGUCHI: Phase III study of bestatin in patients with malignant skin tumors. (1) For malignant melanoma. *Jpn. J. Cancer Chemother.* 12: 77~85, 1985
- OKUYAMA, S. & H. MISHINA: Role of bestatin in the treatment of cancer *via* induction of re-differentiation. Abstracts of 14th Int. Congr. Chemother., S-68-9, p. 235, Kyoto, June 23~28, 1985
- UMEZAWA, H.; T. AOYAGI, S. OHUCHI, A. OKUYAMA, H. SUDA, T. TAKITA, M. HAMADA & T. TAKEUCHI: Arphamenines A and B, new inhibitors of aminopeptidase B, produced by bacteria. *J. Antibiotics* 36: 1572~1575, 1983
- AOYAGI, T.; H. TOBE, F. KOJIMA, M. HAMADA, T. TAKEUCHI & H. UMEZAWA: Amastatin, an inhibitor of aminopeptidase A, produced by actinomycetes. *J. Antibiotics* 31: 636~638, 1978
- AOYAGI, T.; T. YAMAMOTO, K. KOJIRI, F. KOJIMA, M. HAMADA, T. TAKEUCHI & H. UMEZAWA: Forphenicine, an inhibitor of alkaline phosphatase, produced by actinomycetes. *J. Antibiotics* 31: 244~246, 1978
- MORISHIMA, H.; J. YOSHIZAWA, R. USHIJIMA, T. TAKEUCHI & H. UMEZAWA: Synthesis of forphenicinol and forphenicine. *J. Antibiotics* 35: 1500~1506, 1982
- COLLINS, S.J.; R.C. GALLO & R.E. GALLAGHER: Continuous growth and differentiation of human myeloid leukemic cells in suspension culture. *Nature* 270: 347~349, 1977
- COLLINS, S.J.; A. BODNER, R. TING & R. C. GALLO: Induction of morphological and functional differentiation of human promyelocytic leukemia cells (HL-60) by compounds which induce differentiation of murine leukemia cells. *Int. J. Cancer* 25: 213~218, 1980